



Introduction

The ChromoTek Nano-CaptureLigand[™] mouse IgG2a, Fc-specific VHH, biotinylated is used for the sitedirected and specific immobilization of the Fc-fragment of mouse IgG2a in biosensor and ELISA assays. It captures non-biotinylated mouse IgG2a antibodies or Fc-fragments to streptavidin/avidin. Nano-CaptureLigand mouse IgG2a, Fc-specific VHH, biotinylated comprises a monoclonal biotinylated VHH/ Nanobody. The product belongs to the Nano-CaptureLigands[™] family.

Properties

| Description | Monovalent, recombinant single domain antibody for the immobilization of mouse IgG2a: alpaca monoclonal Nanobody, Fc-specific, biotinylated |
|--------------------------------------|--|
| Product Type | Capture Nanobody (VHH) |
| Applications | Immobilization of mouse IgG2a antibodies on avidin and streptavidin surfaces for Bio- Layer Interferometry (BLI), Surface Plasmon Resonance (SPR) and ELISA |
| Target / Specificity | Fc-fragment of mouse IgG2a |
| Cross-reactivity | No cross-reactivity to mouse IgG1, Ig2b, IgG3; human IgG1, IgG2, IgG3, IgG4, IgA; rat IgG1, IgG2a, IgG2b; sera from goat, human, macaque (cynomolgus monkey), sheep Cross-reactivity to mouse IgG2c |
| Affinity (<i>Kd</i>) of monovalent | 1.5 nM |
| (1:1) binding mode | Apparent affinity may be higher for full IgGs due to avidity effects (1 antibody captured by 2 Nanobodies). |
| Concentration | 1 g/L (68 μM) |
| Conjugate | Biotin |
| Degree of biotinylation | On average 1-2 biotin molecules per Nanobody |
| Format | Alpaca single domain antibody, monovalent |
| Host | Alpaca-derived, recombinantly produced in bacteria |
| Clonality | Monoclonal |
| Clone | СТК0108 (VHH0249) |
| RRID | AB_2848188 |



Product code: smsG2aB-1

| Synonyms | Alpaca single domain antibody, VHH, Nanobody, binding domain of single domain antibody, Nano-antibody |
|--------------------|--|
| Validation | Application validated for ELISA and BLI (FortéBio Octet® systems) Determination of cross-reactivity, subclass specificity, sequence, affinity, and melting temperature |
| Purity | Recombinantly expressed and purified via His-tag |
| Form | Buffered aqueous solution |
| Storage buffer | 25 mM TAPS pH 8.5, 500 mM NaCl, 5 mM EDTA, Preservative: 0.09 % sodium azide |
| Storage conditions | Upon receipt store at +4°C/+40°F. <i>Optional</i> : Aliquot upon arrival and store at -20°C/-4°F |
| Stability | Stable for 1 year at +4°C/+40°F |
| Shipment | Shipped at ambient temperature |

Product sizes

| Product | Product code | Size |
|--|---------------|--------|
| Nano Captural izandii mouso IzC2a. Ec specific VIIII, biotipulated | smsG2aB-1-10 | 10 µL |
| Trano-captureLigand — mouse igoza, Ft-specific VHH, biotinyiated | smsG2aB-1-100 | 100 µL |



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Suggested buffer compositions

Recommended buffers for BLI

| Buffer | Composition | |
|---------------------|--|--|
| 1x Kinetics buffer | PBS, 0.01 % (m/v) BSA, 0.002% (v/v) Tween-20 | |
| Regeneration buffer | 0.01 M glycine, pH 2 | |

Recommended buffers for ELISA

| Buffer | Composition | |
|--------------------------------|---------------------------------|--|
| Blocking buffer | PBS, 3 % BSA | |
| Dilution buffer | PBS, 0.05 % Tween-20, 0.5 % BSA | |
| Wash buffer for avidin coating | PBS | |
| Wash buffer | PBS, 0.05 % Tween-20 | |





Bio-Layer Interferometry (BLI) protocol

General notes

- The following protocol was designed for the use with a FortéBio Octet Red96e system. Other BLI systems or your specific research question may require optimization of particular parameters.
- Use the recommended materials or their equivalents.
- Set up all samples in a black 96-well microplate (e.g. Greiner Microplate 96 well, PP, flat-bottom, black, #655209) at room temperature. Use 200 μL per well.
- Nano-CaptureLigands are highly compatible with avidin or streptavidin sensors (e.g. FortéBio Streptavidin (SA) Biosensors, #18-5019) and FortéBio Octet[®] and and BLItz[®] systems.
- Run all experiments at +30°C, a shaking speed of 1000 rpm and a recording rate of 5 Hz.
- Dilute all samples in 1x Kinetics buffer. *Optional:* Use 10x Kinetics buffer.
- NanoCaptureLigands can be regenerated at least 10 times with Regeneration buffer with minimal loss of binding efficiency.
- Nano-CaptureLigands carry a His-tag; thus, avoid the use of anti-His primary antibodies.
- Briefly centrifuge the Nano-CaptureLigand solution before use.

Protocol

1. Baseline 1:

• Incubate the biosensors for 60 s in 1x Kinetics buffer.

2. Loading

- Dilute the Nano-CaptureLigand to a concentration of 1 µg/mL in 200 µL 1x Kinetics buffer.
- Load the diluted Nano-CaptureLigand onto the biosensors for 60-120 s until a loading response of 1 nm is reached.

Optional: Use the threshold limit function in the FortéBio Data Acquisition software.

3. Quenching (optional)

• Incubate the biosensors for 60 s with biocytin (10 μ g/mL in 1x Kinetics buffer) .

4. Baseline 2

• Incubate the biosensors for 120 s in 1x Kinetics buffer.

5. Activation

• Activate the biosensors for 120-180 s with the antibody (20 nM in 1x Kinetics buffer).

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6. Baseline 3

• Incubate the biosensors for 120 s in 1x Kinetics buffer.

7. Association

• Bind different antigen concentrations in 1x Kinetics buffer for 120-600 s. *Note:* As a start, use 0.1-250 μg/mL or 1/10-10x *Kd* of antigen.

8. Dissociation

• Incubate the biosensors for 60-800 s in 1x Kinetics buffer.

Note: Use the wells from step 6, Baseline 3.

Note: Duration of dissociation step depends on the affinity of the analyzed interaction.

9. Regeneration (optional)

- Regenerate the biosensors for 5 s with Regeneration buffer.
- Incubate in 1x Kinetics buffer for neutralization.
- Repeat regeneration 2 times.

Application examples



BLI binding kinetics of a mouse IgG2a anti-V5-tag antibody to a V5-tagged protein. A monoclonal mouse IgG2a anti-V5-tag antibody was immobilized using Nano-CaptureLigand mouse IgG2a, Fc-specific VHH, biotinylated on FortéBio Streptavidin (SA) Biosensors and assayed with different concentrations of a V5-tagged protein.



Sandwich ELISA protocol

General notes

- The following protocol was designed for a standard sandwich ELISA. Other types of ELISA or your specific research question may require optimization of particular parameters.
- Use the recommended materials or their equivalents.
- In this protocol, MaxiSorp plates (e.g. Thermo Scientific[™] White and Black 384-Well Immuno Plates, #460518) are used that must be coated with avidin or streptavidin first. Alternatively, pre-coated avidin/streptavidin plates can be used.
- Nano-CaptureLigands carry a His-tag; thus, avoid the use of anti-His primary antibodies.
- Recommended volumes for 96-well and 384-well microplates:

| Protocol steps | 96-well microplate | 384 -well microplates | |
|--|--------------------|-----------------------|--|
| Coating, antigen binding, antibody binding | 100 µL | 20 µL | |
| Washing, blocking | 300 µL | 90 µL | |

• Briefly centrifuge the Nano-CaptureLigand solution before use.

Protocol

1. Avidin coating (optional)

- Coat each well of a MaxiSorp plate with 10 µg/mL avidin in PBS at +4°C overnight.
- Wash each well twice with PBS.

2. Blocking

- Block each well with Blocking buffer for 1-2 h at room temperature.
- Wash each well 3 times with Wash buffer.

3. Nano-CaptureLigand coating

- Add 50 nM Nano-CaptureLigand (diluted in Dilution buffer) to each well.
- Incubate for 1 h at room temperature.
- Wash each well 5 times with Wash buffer.

4. Immobilization of capture antibody

- Add the capture antibody (diluted in Dilution buffer) to each well.
- Incubate for 1 h at room temperature.
- Wash each well 5 times with Wash buffer.

Note: Test different concentrations of the capture antibody in an initial experiment.

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5. Antigen binding

- Add the antigen to each well.
- Incubate for 1 h at room temperature.
- Wash each well 5 times with Wash buffer.

Note: Test different concentrations of the antigen.

6. Binding of primary antibody

- Add the primary antibody to each well and incubate.
- Wash each well 5 times with Wash buffer.

Note: Dilute and incubate the primary antibody as indicated in the manufacturer's manual.

7. Binding of secondary / detection antibody

- Add the secondary / detection antibody to each well and incubate.
- Wash each well 5 times with Wash buffer.

Note: Dilute and incubate the secondary / detection antibody as indicated in the manufacturer's manual.

8. Detection

- Add the appropriate ELISA substrate solution to each well and incubate as indicated in the manufacturer's manual.
- Analyze with a microplate reader.

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Application examples







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Product overview and related products

| Product | Product code |
|---|--------------------|
| Nano-CaptureLigand™ human IgG/rabbit IgG, Fc-specific VHH, biotinylated | shurbGB-1-10; -100 |
| Nano-CaptureLigand™ human Ig, lambda-LC-specific VHH, biotinylated | shuLB-1-10; -100 |
| Nano-CaptureLigand™ human IgE, VHH, biotinylated | shuEB-1-10; -100 |
| Nano-CaptureLigand™ mouse IgG1, Fc-specific VHH, biotinylated | smsG1B-1-10; -100 |
| Nano-CaptureLigand™ mouse IgG2a, Fc-specific VHH, biotinylated | smsG2aB-1-10; -100 |
| Nano-CaptureLigand™ mouse IgG2b, Fc-specific VHH, biotinylated | smsG2bB-1-10; -100 |
| Nano-CaptureLigand™ mouse IgE, VHH, biotinylated | smsEB-1-10; -100 |
| GFP VHH, biotinylated recombinant binding protein | gtb-250 |

For product details, information, and ordering visit www.chromotek.com.

Product code: smsG2aB-1



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