

Human MMP-1 Sandwich ELISA Kit Datasheet

Please read it entirely before use

Catalogue Number: KE00357

Size: 96T

Sensitivity: 7.3 pg/mL

Range: 19.5-1250 pg/mL

Usage: For the quantitative detection of human MMP-1 concentrations in serum, plasma and cell culture supernatant.

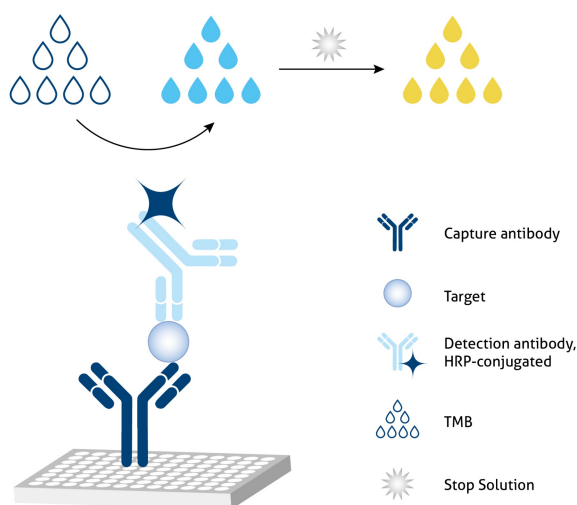
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1. Background

Matrix metalloproteinase 1 (MMP1), which is also known as interstitial collagenase and fibroblast collagenase is a member of the matrix metalloproteinases (MMPs) family. Proteins in this family mainly participate in the breakdown of extracellular matrix both in normal physiological processes and disease processes. Several factors contribute to the expression of MMP1, including endogenous factors, such as polymorphisms and epigenetic regulation in the promoter region of MMP1, as well as exogenous factors, such as the tumor microenvironment. A variety of MMP1 promoter genotypic polymorphisms have been reported in various ethnic populations, and their contributions to different disease risks have been explored. In the tumor microenvironment, a variety of inflammatory factors, including interleukin-8 (IL-8), IL-1 β and tumor necrosis factor- α (TNF- α), have been reported to induce MMP1 expression in cancer cells. (PMID: 31115512, PMID: 28334049)

2. Principle



Sandwich ELISA structure (Detection antibody labeled with HRP)

A capture antibody is pre-coated onto the bottom of wells which binds to analyte of interest. A detection antibody labeled with HRP also binds to the analyte. TMB acts as the HRP substrate and the solution color will change from colorless to blue. A stop solution containing sulfuric acid turns solution yellow. The color intensity is proportional to the quantity of bound protein which is measurable at 450 nm with the correction wavelength set at 630 nm.

3. Required Materials

- 3.1 A microplate reader capable of measuring absorbance at 450 nm with the correction wavelength set at 630 nm.
- 3.2 Calibrated, adjustable precision pipettes and disposable plastic tips. A manifold multi-channel pipette is recommended for large assays.
- 3.3 Plate washer: automated or manual.
- 3.4 Absorbent paper towels.
- 3.5 Glass or plastic tubes to prepare standard and sample dilutions.
- 3.6 Beakers and graduated cylinders.
- 3.7 Log-log or semi-log graph paper or computer and software for ELISA data analysis. A four-parameter logistic (4-PL) curve-fit is recommended.

4. Kit Components and Storage

Microplate - antibody coated 96-well microplate (8 well × 12 strips)	1 plate	Unopened Kit: Store at 2-8°C for 6 months or -20°C for 12 months. Opened Kit: All reagents stored at 2-8°C for 7 days. Please use a new standard for each assay.
Protein standard - 2500 pg/bottle; lyophilized	2 bottles	
Detection antibody, HRP-conjugated (100×) - 120 µL/vial*	1 vial	
Sample Diluent PT 4B1 - 30 mL/bottle	1 bottle	
Detection Diluent - 30 mL/bottle	1 bottle	
Wash Buffer Concentrate (20×) - 30 mL/bottle	1 bottle	
Tetramethylbenzidine Substrate (TMB) - 12 mL/bottle	1 bottle	
Stop Solution - 12 mL/bottle	1 bottle	
Plate Cover Seals	4 pieces	

* Centrifugation immediately before use

5. Safety Notes

- 5.1 Avoid any skin and eye contact with Stop Solution and TMB. In case of contact, wash thoroughly with water.
- 5.2 Do not use the kit after the expiration date.
- 5.3 Do not mix or substitute reagents or materials from other kit lots or other sources.
- 5.4 Be sure to wear protective equipment such as gloves, masks and goggles during the experiment.
- 5.5 When using an automated plate washer, adding a 30 second soak period following the addition of Wash Buffer to improve assay precision

6. Sample Collection and Storage

- 6.1 Serum: Allow blood samples to clot for 30 minutes, followed by centrifugation for 15 minutes at 1000xg. Clear serum can be assayed immediately or aliquoted and stored at -20°C. Avoid repeated freeze-thaw cycles.
- 6.2 Plasma: Use EDTA, heparin, or citrate as an anticoagulant for plasma collection. Centrifuge for 15 minutes at 1000xg within 30 minutes of collection. The plasma can be assayed immediately or aliquoted and stored at -20°C. Avoid repeated freeze-thaw cycles.
- 6.3 Cell Culture Supernatant: Remove particulates by centrifugation for 5 minutes at 500xg and assay immediately or aliquot and store samples at ≤ -20°C. Avoid repeated freeze-thaw cycles.

7. Regent Preparation

7.1 Wash Buffer (1X): If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 30 mL of Wash Buffer Concentrate(20X) to 570 mL deionized or distilled water to prepare 1X Wash Buffer.

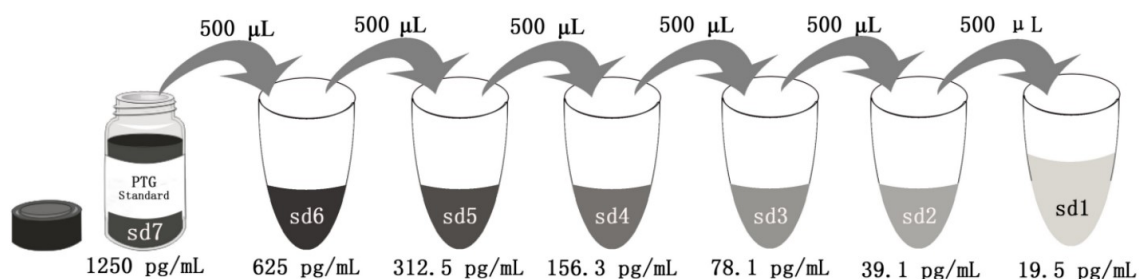
7.2 Detection Antibody, HRP-conjugated(1X): Dilute 100X Detection Antibody, HRP-conjugated 1:100 using Detection Diluent prior to assay. Suggested 1:100 dilution: 10 μ L 100X Detection Antibody, HRP-conjugated + 990 μ L Detection Diluent (Centrifuge the 100X Detection Antibody solution, HRP-conjugated for a few seconds prior to use).

7.3 Sample Dilution: Different samples should be diluted with corresponding Sample Diluent, samples may require further dilution if the readout values are higher than the highest standard OD reading. Variations in sample collection, processing and storage may affect the results of the measurement.

Recommended Dilution for different sample types: 1:16 or 1:32 is recommended for human serum; 1:4 is recommended for human plasma; 1:2 or 1:4 is recommended for cell culture supernatant.

7.4 Standard Serial Dilution:

Add 2 mL Sample Diluent PT 4B1 in protein standard.



Add # μ L of Standard diluted in the previous step	—	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L
# μ L of Sample Diluent PT 4B1	2000 μ L	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L
	"sd7"	"sd6"	"sd5"	"sd4"	"sd3"	"sd2"	"sd1"

8. Assay Procedure Summary

Bring all reagents to room temperature before use (Detection antibody, HRP-conjugated can be used immediately). To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.

8.1 Take out the required number of microplate strips and return excess strips to the foil pouch containing the drying reagent pack and reseal; store at 4°C immediately. Microplate strips should be used in one week.

8.2 Preset the layout of the microplate, including control group, standard group and sample group, add 100 µL of each standard and sample to the appropriate wells. (Make sure sample addition is uninterrupted and completed within 5 to 10 minutes, It is recommended to assay all standards, controls, and samples in duplicate).

8.3 Seal plate with cover seal, pressing it firmly onto top of microwells. Incubate the plate for 2 hours at 37°C.

8.4 Wash

1) Gently remove the cover seal. Discard the liquid from wells by aspirating or decanting. Remove any residual solution by tapping the plate a few times on fresh paper towels.

2) Wash 4 times with 1X Wash Buffer, using at least 350-400 µL per well. Following the last wash, firmly tap plates on fresh towels 10 times to remove residual Wash Buffer. Avoid getting any towel fibers in the wells or wells drying out completely.

8.5 Add 100 µL of 1X Detection antibody, HRP-conjugated solution (refer to Reagent Preparation 7.2) to each well. Seal plate with cover seal and incubate for 40 minutes at 37°C.

8.6 Repeat wash step in 8.4.

8.7 Signal development: Add 100 µL of TMB substrate solution to each well, protected from light. Incubate for 15 to 20 minutes. Substrate Solution should remain colorless until added to the plate.

8.8 Quenching color development: Add 100 µL of Stop Solution to each well in the same order as addition of the TMB substrate. Mix by tapping the side of the plate gently. NB: Avoid skin and eye contact with the Stop solution.

8.9 Read results: Immediately after adding Stop solution read the absorbance on a microplate reader at a wavelength of 450 nm. If possible, perform a double wavelength readout (450 nm and 630 nm).

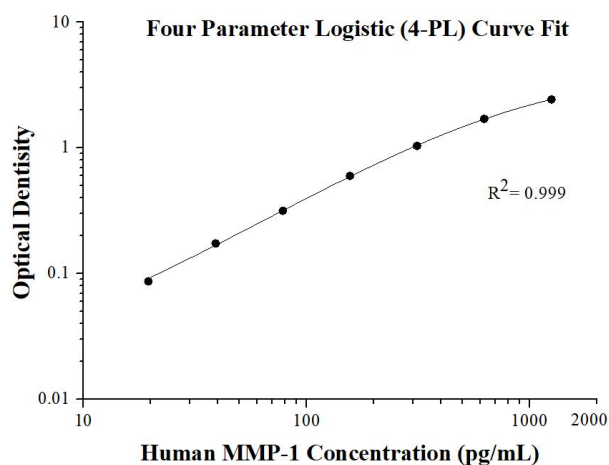
8.10 Data analysis: Calculate the average of the duplicate readings (OD value) for each standard and sample, and subtract the average of the zero standard absorbance. Construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis, use four-parameter logistic curve-fit (4-PL) analysis to do this. If the samples have been diluted, the OD readout from the standard curve must be multiplied by the dilution factor used.

Step	Reagent	Volume	Incubation	Wash	Notes
1	Standard and Samples	100 µL	120 min	4 times	Cover Wells incubate at 37°C
2	Diluent Detection antibody, HRP-conjugated Solution	100 µL	40 min	4 times	Cover Wells incubate at 37°C
3	TMB Substrate	100 µL	15-20 min	Do not wash	Incubate in the dark at 37°C
4	Stop Solution	100 µL	0 min	Do not wash	-
5	Read plate at 450 nm and 630 nm immediately after adding Stop solution. DO NOT exceed 5 minutes.				

9. Validation Data

9.1 Standard curve

These standard curves are provided for demonstration only. A standard curve should be generated for each set of samples assayed.



(pg/mL)	O.D	Average	Corrected
0	0.0341 0.0359	0.035	-
19.5	0.123 0.1207	0.1219	0.0869
39.1	0.2138 0.2045	0.2092	0.1742
78.1	0.3599 0.3428	0.3514	0.3164
156.3	0.6495 0.6197	0.6346	0.5996
312.5	1.0936 1.0574	1.0755	1.0405
625	1.7785 1.7034	1.741	1.706
1250	2.4818 2.4534	2.4676	2.4326

9.2 Precision

Intra-assay Precision (Precision within an assay) Three samples of known concentration were tested 20 times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays) Three samples of known concentration were tested in 24 separate assays to assess inter-assay precision.

Intra-assay Precision				
Sample	n	Mean (pg/mL)	SD	CV%
1	20	950.1	24.4	2.6
2	20	205.6	5.2	2.5
3	20	50.3	2.1	4.1

Inter-assay Precision				
Sample	n	Mean (pg/mL)	SD	CV%
1	24	947.1	62.1	6.6
2	24	201.9	4.3	2.1
3	24	53.9	4.8	9.0

9.3 Recovery

The recovery of human MMP-1 spiked to three different levels throughout the range of the assay in various matrices was evaluated.

Sample Type		Average% of Expected	Range (%)
Human serum	1:128	92	86-108
	1:256	89	77-101
Human plasma	1:16	87	86-89
	1:32	83	79-85
Cell culture supernatant	1:64	101	72-125
	1:128	106	72-124

9.4 Sample values

Human serum/plasma - human serum and plasma samples were evaluated for the presence of human MMP-1 in this assay.

Sample Type	Mean (pg/mL)	Range (pg/mL)
Human serum (n=16)	9,124.3	3,045.5-19,864.3
Human plasma (n=8)	653.4	99.3-1,631.8

Cell culture supernatant - HT-1080 were cultured in DMEM supplemented with 10% fetal bovine serum, 100 U/mL penicillin and 100 µg/mL streptomycin sulfate. Aliquots of the cell culture supernate were removed and assayed for levels of human MMP-1, and measured 907.5 pg/mL.

9.5 Sensitivity

The minimum detectable dose of human MMP-1 is 7.3 pg/mL. This was determined by adding two standard deviations to the concentration corresponding to the mean O.D. of 20 zero standard replicates.

9.6 Linearity

To assess the linearity of the assay, samples were diluted with the appropriate **Sample Diluent** to produce samples with values within the dynamic range of the assay.

(The human serum samples were initially diluted 1:16. The human plasma samples were initially diluted 1:2. The cell culture supernatant samples were initially diluted 1:8.)

		Human serum	Human plasma	Cell culture supernatant
1:2	Average% of Expected	100	100	100
	Range (%)	-	-	-
1:4	Average% of Expected	104	109	107
	Range (%)	96-113	101-119	106-108
1:8	Average% of Expected	99	116	104
	Range (%)	85-114	108-121	98-108
1:16	Average% of Expected	96	113	103
	Range (%)	77-121	105-124	85-114

9.7 Specificity

This assay recognizes natural and recombinant human MMP-1.

The following factors prepared at 50 ng/mL were assayed and exhibited no cross-reactivity or interference.

Recombinant human:

MMP-3

MMP-9

TIMP-1

TIMP-2

A sample containing 50 ng/mL of the recombinant human MMP-2 read as 64.53 pg/mL (0.129% cross-reactivity).

10. References

1. Shen CJ, Kuo YL, Chen CC, Chen MJ, Cheng YM. MMP1 expression is activated by Slug and enhances multi-drug resistance (MDR) in breast cancer. PLoS One. 2017 Mar 23;12(3):e0174487.
2. Chen Y, Peng S, Cen H, Lin Y, Huang C, Chen Y, Shan H, Su Y, Zeng L. MicroRNA hsa-miR-623 directly suppresses MMP1 and attenuates IL-8-induced metastasis in pancreatic cancer. Int J Oncol. 2019 Jul;55(1):142-156.
3. Young-Min SA, Beeton C, Loughton R, Plumpton T, Bartram S, Murphy G, Black C, Cawston TE. Serum TIMP-1, TIMP-2, and MMP-1 in patients with systemic sclerosis, primary Raynaud's phenomenon, and in normal controls. Ann Rheum Dis. 2001 Sep;60(9):846-51.
4. Murawaki Y, Ikuta Y, Idobe Y, Kawasaki H. Serum matrix metalloproteinase-1 in patients with chronic viral hepatitis. J Gastroenterol Hepatol. 1999 Feb;14(2):138-45.