CoraLite® 488-594 Phalloidin

Stock preparation

Prepare a 200T/mL stock solution by dissolving lyophilized phalloidin (300T) in 1.5mL sterile water and protect solution from light.

Fluorescence microscopy

1. Wash the cells 3 times with PBS.
2. Fix the cells on ice for 15 min with 4% paraformaldedehdyde in PBS.
   Note: Avoid fixatives using methanol because they can destroy actin during the fixation process.
3. Wash the cells 3 times with PBS.
4. Permeabilize the cells with 0.2% Triton X-100 in PBS at room temperature for 5 min.
5. Wash the cells 3 times with PBS.
6. Prepare master mix by diluting 1-5μL working stock solution with 200μL PBS. For multiple samples, scale appropriately.
7. Add 200 μL master mix to each sample, and incubate for 20 min at room temperature for staining. Protect from light.
   Note: To avoid evaporation of the dye solution during the incubation, place samples in a humid environment.
8. Wash the cells 2-3 times with PBS.
9. Observe the samples under a fluorescence microscope.